parallel testing was performed using Chemfort VA from which the filter system had been removed.

Results No drug was found in any of the test samples with the intact air filter system in Chemfort VAs, either fresh, following aging for 3 years or after 7 days of exposure to drug vapours. Recovered vapour was consistently found in the positive control samples which had Chemfort VAs without a filter system. Mean±SD (n=5) levels were 69±34 and 35±20 ng for cyclophosphamide and 5-FU, respectively.

Conclusion and relevance The results confirm the efficacy of the Chemfort air filtration system, even after 7 days of exposure to drug vapour or a shelf life of 3 years.

REFERENCES AND/OR ACKNOWLEDGEMENTS

Conflict of interest Corporate sponsored research or other substantive relationships:

The author is an employee of Simplivia Healthcare.

3PC-012 MANAGEMENT OF NON-ADMINISTERED CHEMOTHERAPY PREPARATION: AN OPPORTUNITY TO MINIMISE DRUG WASTE IN ONCOLOGY PHARMACY

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Background and importance The non-administration of cytotoxic preparations contributes significantly to drug waste and costs in the centralised cytotoxic preparation units (CCPU). Monitoring and proper management of returns of preparations could reduce drug wastage.

Aim and objectives The aim of the study was to analyse the reasons for returns and quantify reused cytotoxic preparations before and after implementation of corrective measures.

Material and methods A prospective study was conducted at our hospital pharmacy at the National Institute of Oncology over two 8 month periods (January to August 2018, January to August 2019). Data on the reasons, content and fate of returns were collected and analysed.

Results At the end of the first period, 125 preparations corresponding to 90 prescriptions were returned. Absence of the patient was the most common reason (56%), followed by crystallisation of product (19%), mainly taxanes. Docetaxel was the most returned preparation (17.6%). The corrective measures taken were: optimisation of communication between the CCPU and clinical services, strict dilution of taxanes and etoposide in glass vials and updating of physicochemical and microbiological stability sheets for cytotoxics. During the two study periods, we found a similar number of returns (0.6%) corresponding to 15 851€ and 16 874 €. The absence of the patient, the most frequent reason in the two periods, decreased from 56% to 40%. Product crystallisation decreased considerably (19% vs 2%). The number of reasigned preparations increased from 2.4% to 7%. Reusing corresponded to 64€ and 2760€ for period 1 and period 2, respectively.

Conclusion and relevance This study found a high number of preparations returned due to crystallisation by taxanes via interactions containing content. Updating the stability data of the anticancer drugs used in our hospital based on recent

international guidelines and follow-up of chemotherapy preparations had a significant impact on the reasons and cost of returns. Vigilance by pharmacists is required when validating prescriptions in order to minimise the avoidable causes of chemotherapy wastage and to make savings.

REFERENCES AND/OR ACKNOWLEDGEMENTS

No conflict of interest.

3PC-013 COMPARATIVE BIOPHYSICAL STABILITY STUDY OF ZIV-AFLIBERCEPT (ZALTRAP, OPENED VIALS) STORED AT 4°C AND ROOM TEMPERATURE FOR 2 WEEKS

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Background and importance Ziv-aflibercept (Zaltrap) is an Fc-fusion protein used in the treatment of colorectal cancer. Changes in the structure or aggregation, which may arise from handling and storage, may affect the efficacy of the treatment and it could cause severe immune reactions in patients. The shelf life indicated by the manufacturer for the unopened vial is 3 years; there is no information on the surplus of opened vials.

Aim and objectives To compare the biophysical stability of ziv-aflibercept (Zaltrap) stored refrigerated at 4°C and at room temperature protected from light for 2 weeks.

Material and methods Three independent samples of fresh ziv-aflibercept were collected from hospital and stored in amber glass vials protected from light at 4°C and at room temperature.

Particulate: dynamic light scattering (DLS) readings were carried out in a protein solution DynaPro-99 system dynamic light scattering module equipped with a temperature control micro sampler (Wyatt, Santa Bárbara, California, USA) for obtaining the hydrodynamic radius and polydispersity.

Tertiary structure: intrinsic tryptophan fluorescence measurements were carried out on a Cary eclipse spectrofluorometer (Agilent, Santa Clara, California, USA). Each spectrum was reduced to a single a dimensional number (centroid):

$$C = \frac{\sum_{i=1}^n (f_i \lambda_i)}{\sum_{i=1}^n f_i}$$

LMW aggregates: size exclusion chromatography (SEC) was used. The analysis was performed by liquid chromatography using an Agilent 1100 chromatograph equipped with a quaternary pump, degasser, autosampler, column oven and photodiode array detector (Agilent).

Results No significant changes were detected in the samples stored refrigerated by any of the techniques used: aggregation did not occur, supported by the results from DLS and SEC. No changes in conformation were detected: fluorescence centroid was maintained.

Significant changes were detected in the samples stored at room temperature: the start of aggregation was detected by SEC but larger aggregates were not detected by DLS. Centroid value increased significantly, indicating conformational modifications.